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Spring 2021



SCARLET ELF CUP

Sarcoscypha austriaca

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View from the Chair

(This is a transcript of Irene's report to the AGM)

This is going to be a very brief report because little happened last year and because we are on Zoom we want to keep things as brief as possible. It will also be my last report as Chair so I'll begin with:

The **Committee**: Firstly, Jeanette Maddy, who has been Vice-Chair and did a great job when I was ill as well as organising the DNA sequencing group and acting as Zoom host, is willing to take over as Chair. The Group couldn't be in better hands and I hope you support this change.

Secondly John Lovelady has resigned from the committee, so thank you John for your past work and I have agreed to take over his role as Outreach Co-ordinator. Thirdly, we welcome onto the Committee two new members Kirsten Nagel and Helen Speed. And finally, Max Furmidge has taken over as manager of our website.

All other committee members remain and continue with their current jobs. So I hope you will be happy with these proposed changes and the membership when voting for the committee and officers later in this meeting.

Apart from the 2020 AGM the Covid pandemic has meant that no forays or workshops could take place last year except for a few forays restricted to 6 people in late August to early September when restrictions eased. John Watt will report on these in the next Newsletter. But Newsletters have been produced, we've had committee meetings via Zoom and the email group and Facebook page have continued to flourish. So many thanks to all who made this possible and especially Paul Hamlyn and Mike Walton for the Newsletter, Tony Carter for the email group and Lynne Healey for the Facebook page. We very much hope that forays, including the Keswick residential weekend, and workshops, including a Microscope workshop, can be resumed in 2021 but no firm dates can be given until the proposed easing of lockdown restrictions actually happens. So as soon as we can, we will email you about this.

However, we felt that something extra should be done for members so we are producing online help with fungal identification, currently in the form of Zoom PowerPoint presentations. I've done the first one, which is on *Mycena*, the Bonnet fungi. It's intended chiefly to help beginners and anyone who wants to improve or brush up their i.d. skills, and we hope this can go out during March. So watch out for an email and link about this and also about other presentations later.

All being well, Jeanette will produce one on waxcaps and the hope is that presentations can be recorded and made available to people who couldn't watch on the original date – although, of course, people then couldn't ask questions and take part in discussions.

That's all I want to report but do feel free to ask questions and, finally I just want to say how much I've enjoyed all my years as Chair of North West fungus Group. It's been a privilege.

Irene

Editorial

After a year of lockdowns there does at last appear to be a glimmer of light at the end of the tunnel. Fortunately, some members have been able to report their favourite finds from sole forays for the newsletter and I myself discovered a local woodland just down the road from where I live that I had not paid much attention to in the past. John Watt has completed his recorders report for 2020 with details of the fourth miniforay carried out last year. The article by Tim Rogers on Ainsdale must be a record for the number of photographs included and thanks to Jeanette for a fungal quiz.

Some other good news reported at the NWFG AGM Zoom meeting is that our President, Geoffrey Kibby announced that Volume 3 of Mushrooms and Toad-stools of Britain & Europe would be published later this year.

Many thanks to all those members who have contributed articles for this issue, and to Mike Walton for typesetting and sending out the newsletter. Articles can be submitted to me by email. Pictures of fungi to accompany articles are very welcome preferably sent as separate attachments. Please note that it is important to show due diligence when including any photographs (or other material) that have not been taken by yourself by getting permission and including the name of the photographer (or copyright holder) so that due credit can be given in the newsletter.

Paul F. Hamlyn pfhamlyn@gmail.com

North West Fungus Group - Spring 2021

My favourite find of 2020.

In November 2021, I was leading a group of University students around Scutchers Acres for their Geographical Information System Phase1 Habitat survey mapping exercise. They were pleased to be told the names of plants or fungi we encountered but I was stumped (sic) when we came across three Little Brown Mushrooms (LBM) growing on a rotten stump. Much smaller than *Pluteus*, they had a 15mm smooth hemispherical brown cap and beige-coloured adnexed gills, and a concolourous central stipe.

The overnight spore print was brown - closely matching number 14, rusty-tawny, on the British Mycological Society colour chart ruling out any of the *Hypholoma*, *Pholiota* or

Psathyrella genera. Next step anyway was to look at the spores themselves, and these were not only totally smooth, lacked either a germ pore or an apiculus (mostly), and many were phaseoliform (bean-shaped) making them look quite distinctive. They measured (7) 8.1 (9) x (4.5) 4.9 (5.5) μ , Q = 1.63. (Below right)

I thought these features would be enough to test the diagnostics of Mycokey 4.1 which only now works on my old lap-top; but this didn't come up with a genus. The Laessøe and Petersen LBM wheel key displays 20 different small genera, but none had the distinctive spore shape.

(I admit I don't often use the *Funga Nordica* Genus key, because it's quite hard work, but may have been worth considering.)



Pileocystidia were typically 110 μ x 12.5 μ , cylindical. (Opposite)

I then turned to the wonderful illustrations by Ludwig in his *Pilzkompendium* volume I, actually just flicking the pages from the back, looking especially for





lignicolous species, upon which I reached the genus *Simocybe* which instantly made sense. In addition to the pleurotoid (lacking stem) *S. haustellaris*, those with stems are basically those two growing on herbaceous substrate and the two on rotten wood, *S. centunculus* (and its varieties) and *S. sumptuosa*, both quite unusual especially the former with only four records on old FRDBI. Bon's Field Guide does include the former as representative of the genus with its characteristic spore. The species are also in Buzcazki and Eysssartier books.



The field characters of these two are rather similar and microscopic features are also close.

	Simocybe sumptuosa	<i>Simocybe</i> in question	Simocybe centunculus
Basidia	2 or 4 spored	2 or 4 spored	4 spored
Spores	6-12 x 4-7 (Ludwig) 8-9.5 x 5-5.5 Q = 1.5-1.8 (FN)	8.1 x 4.9 Q = 1.63	6-8.5 x 4.5 -5.5 (Lud- wig) 6-8.5 x 4-6 Q = 1.4 - 1.7 (FN)
Cheilocystidia	clearly capitate heads over 8µ	Capitate head to 10µ	Some capitate up to 8µ across
Pileocystidia	Cylindrical; => 80µ	Cylindrical; 110µ	Pear or bottle shaped, shorter.

Using the Ludwig and FN keys, the likely species is *Simocybe sumptuosa*:

With 231 records on FRDBI.info *S. sumptuosa* is less rare than *S. centunculus* though there are only three were recorded north of the midlands, so I am really happy with the find, and I may now keep this genus in mind for the future. It is also one on which could be worth running a DNA analysis so it has been dried for posterity.

John Watt

2022 A.G.M.

Saturday 26 February 2022 at 10.30 a.m. (10.00 a.m. Coffee)

Risley Moss Centre.

Stenner Woods

Paul F. Hamlyn

Adjacent to Fletcher Moss Park in Didsbury, a suburb of Manchester, Stenner Woods is a small area of wet woodland a remnant of the original Mersey Valley flood plain. Fallen trees and dead wood are generally not removed from the site providing a habitat for invertebrates that in turn provide a food source for birds. This also makes the site an ideal location for wood decomposing fungi.

With restrictions on travel and attending forays in 2020 I have started to record at this site since it is only a short distance from where I live. I have only found common species so far but even these sometimes require careful examination.



Ganoderma applanatum

Spore size measurements using Piximètre

For example, *Ganoderma applanatum* (Artist's Bracket or Artist's Conk) is similar in appearance to *Ganoderma australe* (Southern Bracket) but there is a significant difference in spore size (Andy Overall, Field Mycol. 17(4):124-128, October 2016). Measuring spores is tedious at the best of times but there are free software programs that can help. Robin Dean recommends Piximètre so I put it to the test and it worked very well.

One day I came across a rather soggy and partially eaten specimen of *Pluteus cervinus* (Deer Shield - opposite). It appeared to have the requisite free and pale pink gills. A quick look under the microscope revealed the large cystidia on the gill edges with horn-like prongs characteristic of this species.



Pluteus cervinus Cheilocystidia (right)



Sarcoscypha austriaca Excipular hairs (right)



In January 2021 the River Mersey came within centimetres of the top of the riverbank necessitating use of the overflow water storage basin that includes Stenner Woods. Within a few days most of the water had receded and it was safe to walk around the woods again. At the beginning of February, I was greeted with an abundance of *Sarcoscypha austriaca* (Scarlet Elfcup), they obviously like very wet conditions. *Sarcoscypha coccinea* (Ruby Elfcup) is almost identical to *S. austriaca* but has straight hairs on the outer (infertile) surface of the cups whereas the excipular hairs of *S. austriaca* are strongly sinuous to coiled and convoluted (Butterfill & Spooner, Mycologist 9 (1):20-26, 1995).

Note: all photographs taken by Paul F. Hamlyn.

North West Fungus Group - Spring 2021

Recorders report for 2020

It was only during the late summer when, under the Rule of 6 for outdoor meetings, the group could stage a few miniforays, and three of these have been reported upon in the previous newsletter.

The other fourth miniforay was led by Penny Hinsiger and Mike Walton on 20 September at Finsthwaite Heights, a venue new to the group.

It is a delightful site with natural Oak woodland on the approach up to the dam around where it is more damp with correspondingly more fungi than the drier slopes on the approach. Up there the predominant tree was Birch (*Betula*) but Aspen (*Populus tremula*), Alder (*Alnus glutinosa*), Hazel (*Corylus avellana*) were also plentiful but in addition there had also been plantings of Larch (*Larix*) and Pine (*Pinus*), so all these offered a wide range of hosts for their mycorrhizal associates.

Of particular interest was the finding of four species of *Leccinum*, two of which with slight blueing reactions were unfamiliar to the forayers, requiring further homework including the examination of cells in the cap cuticle. The *Leccinum schistophilum* is a *Betula* associate previously found by Peter Smith of NWFG at Longworth Clough, but with only 33 records nationally. The *Leccinum cyanoba-sileucum* is a northern species with 253 records and also associated with *Betula*, and has been recorded by Bruce Ing at Cliburn Moss. *Leccinum scabrum* is the one associated with *Betula* which we are most familiar with and by comparison has 2,600 records nationally. The fourth species in this genus we found, growing under *Populus tremula*, was *Leccinum duriusculum* which has 66 records nationally but has often been found by Tony Carter around his golf course, and John Watt at Scutchers Acres.

Of the dozen odd species of *Leccinum* in this country, seven are *Betula* associates; two with *Populus*; two with *Quercus* including *Q ilex;* one with *Carpinus / Corylus;* one with *Pinus;* whilst in contrast *Leccinum aurantiacum* is versatile and associates with a wide range of tree species and is reasonably common.

The *Hebeloma* genus was also well represented with *H. velutipes* and *fragilipes* but another species had Irene working on it for a long time, measuring the dimensions, and their relative ratios, of many cystidia. The most likely species was *H. ingratum*, which has just two records nationally.

Melanoleuca polioleuca (from the Greek black-white, grey-white) when found on site indeed had much a more grey cap than one expects but that was because it was very dry and on becoming more moist had much richer brown sepia tones. Though common enough, the observation of septate cystidia can help towards this confirmation. (Photos - top right)





One of my favorites was the Snaketongue Truffleclub, *Tolypocladium ophioglossoides - (right)* to give it the current scientific name - though we couldn't find the buried *Elaphomyces* truffle host. The other was *Laccaria bicolor*, Bicoloured Deceiver, (below) which exhibited lovely subtle textures and lilac colouration towards the base of the stipe.



The total tally for the day was 50 species and we hope we may be able to revisit this site to amplify these records.

John Watt 10/2/2021

T. ophioglossoides

The information

FAVOURITE LOCKDOWN FINDS

Kathleen Ryan

Almost every day I walk on Billinge Hill in Blackburn which is close to where I live. The part of the hillside where I walk is mixed woodland with some old beeches and Scots pine and one or two exotic species e.g. Monkey Puzzle trees, planted in the 19th century when it was part of the Fielden estate. It is a public space and heavily used by dog walkers, runners and mountain bikers.

Each year I find a variety of fungi, though I have only ever found common species. The year 2020 seemed to be a poor year for finds. Usually there are plenty of the common *Russula* species – *ochroleuca, fellea, betularum, mairei, vesca* – and a few of my favourite, the Yellow swamp russula (*R. claroflava*). This year there were hardly any russulas, although the Tawny grisette (*Amanita fulva*) was everywhere.

If I walk to the nearest postbox I pass an urban grass verge planted with hornbeams. One small part produces *Amanitas - rubescens, muscaria*, occasionally *spissa*, several *Tricholomas* and *Cortinarius* species and the occasional *Boletus edulis*. In early August I was excited, to see a Death cap (*Amanita phalloides* right). A very typical specimen with a streaky brownish olive cap. It just seemed so unexpected in such an urban situation.





In December when I was no longer expecting to see anything at all, there shining in the sunshine in the wood, were clusters of what I assumed to be the Glistering inkcap (*Coprinellus micaceus* left) on a small dying birch tree at the edge of the path.

After a wet night no flecks of the 'mica' were visible initially, but moving a leaf confirmed what I thought.

So, whilst my 2020 year

may not have been exceptional for fungi, what I found certainly provided me with surprise and interest even if unable to join others on forays.



LEPISTA PERSONATA

Tony Carter

In September 2019 I was foraying in Calderstones Park, Liverpool. Passing a small mixed plantation, I noticed a group of fungi under a cluster of yew trees. I had to crawl underneath the trees to collect a specimen for examination. Having crawled out again, I saw they had a light coloured cap and pale mauve stem. My immediate conclusion was *Lepista saeva* now *personata*, the Field Blewit.

I took it home to check because, although most of the area is open grass, these were clearly associated with yew. I consulted the British Database and British and Irish Checklist. This fungus is found 'On soil amongst grass in scrub woodland with deciduous trees and shrubs. Also known on heathland, in unimproved grassland and under planted trees in roadsides and gardens'. Association with yew or any conifer is not mentioned.



I began to doubt my identification, perhaps it was *Lepista irina* or a pale *sordida*. I posted a photograph of my specimen to the Email Group. The replies were all – *Lepista personata*.

In December I was at my local golf course when I noticed a group of fungi under some *Cupressus leylandii* in a small mixed plantation surrounded by grassland. Exactly the same circum-

stances as September. Again, I had to crawl on hands and knees under a tree to extract a specimen. Again, l identified *Lepista personata*.

This specimen I dried and took to the DNA workshop at Risley where I extracted the DNA and submitted the sequence to the relevant databases on the internet. The results indicate that my DNA sequence matches other sequences that have been submitted as *Lepista saeva/personata*.

A month later, I was back on the golf course at an area normally off limits except that golf was terminated due to lockdown. In another small mixed copse, tucked away under some *Cupressus leylandii* was another group of *Lepista personata* and yet another crawl to collect and identify.

Both park and golf course are areas dominated by maintained grassland. Both have small isolated mixed copses that include a tight cluster of conifer under which these Lepistas grow in sheltered conditions. This association with conifer is too consistent to be chance and yet not previously recorded.

Book Review

21st Century Guidebook to Fungi, 2nd Ed. David Moore, Geoffrey Robinson, Anthony Trinci.

The study of biology at school and indeed even at university deals overwhelmingly with plants and animals, the world of fungi being the Forgotten Kingdom. However, this distillation of their lifetimes' study of the world of fungi by these authors, in this second edition, reveals both the fundamental importance of fungi on our planet and at the same time an astonishing existing depth of knowledge of the biology of fungi in all facets.

As I developed my interest in field mycology, I was intrigued by the many mysteries of the fungal lifestyle so was delighted to find a second hand copy of the 1961 classic *Biology of Fungi* by Ingold. At another BMS autumn Kew meeting I purchased a second hand copy



of Carlile and Watkinson's 1994 *The Fungi*. Still later, I acquired the 2001 *Introduction to Fungi* by Webster and Weber. I am glad that I then awaited the appearance of this second edition of *21st Century Guidebook to Fungi* by our own Manchester team of academics.

With nineteen Chapters over 600 pages the authors take one along so many different avenues. One could perhaps read a chapter out of sequence with excellent cross-references, but I was instantly drawn to the early chapter on the evolutionary story of fungi and how they facilitated colonisation of the land in the earliest of times. I loved the sci-fi illustrations of the nine metre high Prototaxites, a likely fungus from 400 m years ago, but there are several different fungal phyla represented in the Rhynie Chert fossils from Aberdeenshire from the same period.

The following chapter is a nicely illustrated review of the natural classification of fungi. This is systematically set out in Appendix 1, a very useful up-to-date quick source of reference. Within this chapter there is a discourse on the 'Species concept' as applied to fungi. The morphologically based approach clearly fails. So also does the biological approach for many fungi cannot be mated in the laboratory or are usually in an asexual state. The ecologically based approach is

largely limited to plant pathogens and even here has limitations, so the phylogenetic, or evolutionary, approach based on DNA analysis has most promise.

Genetics is dealt with at some length but despite the complexity, key concepts are nicely tabulated, so that for instance the so-called ITS region (internally transcribed spacers) of the non-structural rRNA is a variable domain allowing characterisation of fungi to species level. Four other different areas of rRNA enable characterisation to higher or lower levels of taxonomy, for instance to phylum or to race.

Fungal reproduction is bewilderingly diverse and complicated and variable even within a phylum. Some fungi reproduce asexually by conidiogenesis which may give them a competitive advantage on wastefully energetically expensive reproduction machinery. Sexual reproduction strategies range from self-fertile homothallic species to self-sterile species with two or more mating types. The classic extreme case is of *Schizophyllum commune* (Common Porecrust) with 288 possible mating types, which serves to ensure a high chance of successful outbreeding.

Whereas much hyphal physiology is known, there still remain many mysteries underpinning hyphal behaviour, such as the signalling to initiate hyphal differentiation. Several useful pages with helpful drawing are devoted to the "10 ways to make a mushroom". To complement morphology at the microscopic level, Appendix 2 proceeds through illustrations of the many different types of conidiogenesis; the different hyphal types as one may encounter in resupinates and brackets; and I especially liked the clearly laid out and named different cystidial types, my point of reference for the future.

Another chapter deals with fungi as symbionts and predators of animals, in which latter category include the 700 or so fungal species, in four main groups, which have independently developed different strategies for the killing of nematodes. The pathogenicity of fungi in plants and animals are discussed extensively, and subsequently the antifungal strategies are reviewed.

Then there are wide-ranging accounts of how fungi are exploited as food, including the 'gardening' termites and ants. Finally, the biotechnology of the fungal industry, from traditional low-tech to high tech, includes a six page story of the development of Quorn \mathbb{R} (*Fusarium venenatum*) from laboratory to the supermarket.

So, whilst this is an academic textbook - with frequent recommendations for further reading - there are answers to nearly every question you may have when next you find yourself musing on a particular toadstool sprouting magically – how it got there, why it likes it there or why it smells in a particular way.

John Watt

Ainsdale

Tim Rogers

Since moving to Formby late last year I'm ideally placed for Ainsdale, the Dune Heath, Ravenmeols NR etc., – this started as a description of a single foray, but I've accumulated so many photos over 2020, it seems a shame to limit myself to this – so here we go.

The original foray was through the mixed woodland (mostly *Pinus & Betula*), in part of the NNR you won't have seen, between the Fisherman's Path, and the area we visit with Tony - nothing unusual, but have chosen photogenic species, mostly ones you'll be familiar with; Clubfoot, fairly common, but very striking; Bloody Brittlegill (*Russula sanguinea* – under *Pinus* - right); three in one (below), Summer Bonnet (*Mycena abramsii*), with *Lycoperdon pyriforme*, & *Mycena galopus* var. *nigra* (on Salix); *Stereum subtomentosum* (on Betula). There were many more species, of course



(this was during one of several excellent 'flushes'), but these produced the best photos!



A similar walk, a month later (November), produced, in particular, two Tricholomas – *T. scalpturatum* (under *Quercus & Betula - below*), &, locally,



in enormous numbers, under Betula, *T. Stiparophyllum* (right), well worthy of its name of Chemical Knight (intensely aromatic, but not unpleasant - to me).

I've included *Lepista nuda* (now *Clitocybe nuda* - below), this one found on the Dune grasslands, because of the wonderful intensity of its colour.



A previous walk over the dunes produced what I took to be *Geastrum rufescens*



(below), but, as you will see, it appears to have a halo, which it shouldn't – but nothing else seems to fit.





My walks started in May, when the best finds were *Cerrena unicolor* (above), and *Hypholoma ericaeoides* (right), both far from common. Later in the summer, at Larkhill Fields, I found *Lycoperdon utriforme* (right) – very distinctive.

Many of you will know the Dune Heath, which is much underrated by some, but has produced many excellent records – a recent one is *Clavaria argillacea* (Moor Club), which occurs very late, but in some abundance – it only grows with *Calluna* (common heather), apparently (another reason for it being largely unknown to our Group);



I've also included *Hygrophorus hypothejus* (Herald of Winter - right), which appeared at the same time, and is a particular favourite.





A very characteristic species of the Dunes, but only occurring in the NNR in tiny numbers (locally abundant elsewhere), is *Tulostoma brumale* – two excellent examples included here.



Another Dune grassland species which is (according to Tony) much recorded (though not by me), is *Lepiota erminea* – these (below) were also found rather late, well into November.





Melanoleuca species are widespread locally – the one included here is *melaleuca* – right - (no cystidia found, after very thorough search), a species which is 'often recorded, but poorly known' (CBIB) - most preserved material is apparently *M. Polioleuca*;



Polioleuca; from Funga Nordica & Collins field guide. Some of you may find the inclusion of Auricularia auricula-



judae (Jelly Ear) a bit strange, but these are very fresh specimens, & very pleasing, showing their 'veins', which are not often so apparent.

Couldn't finish without mentioning *Cortinarius* – have found *flexipes*, *hemitrichus*, and *obtusus*, but included here is *C*. *umbrinolens*, with its characteristically veined cap (see B&K), found at Ravenmeols, under *Betula*.





And, finally, a quiz from Jeanette to while away lock-down.

Answers from mike.a.walton@googlemail.com

Mushrooms, but not Toadstools i.e the missing letters can be found in the word 'toadstools'		
1. FE _EHC_P		
2HEHE_ WU	F_	
3RY'E		
4U_PHUR _UF_		
5. YEWG_H_R	N	
6ER_YING _NGE_	-	
7N_YIBA		
8. GEN _PIN_E_		
9 E V I _ ' H		
10. F_Y _G_RIC		
11H_ P_R		
12. CRE_ E_R_H_	R	
13. PE _YER		
14C_R_E_IN_ BE	_E	
15. GEN BEG		
16I_RY _M_NI		
17YC_M_RER_P	_	
18PE_ BRIE_	_ E M	
19. W WYF		
20HE_ CRU		
warnes from BWS lists of English names		

North West Fungus Group - Spring 2021

Wanted

Collections of Coprinopsis pseudoradiata.

I am on the lookout for collections identified as *Coprinopsis pseudoradiata*. There are surprisingly few deposited collections from the UK despite a reasonable number of records on FRDBI. Several identified as this species (including one by me in 1999!) have turned out to be the more recently described *C. candidolanata*.

These are rather small species growing on dung of various animals with veil shaped like strings of sausages (with or without additional diverticulate veil). The spores are small (around 9 x 5 μ m). *C radiata* is much more common and has longer spores (usually >12.5 on average) and *C. cinerea* is also very common, especially on dung/straw heaps and has somewhat larger spores (9-11 x 6-7 μ m), but also a larger, more robust fruitbody with a firm base to its stipe. The fruitbodies turn to ink fairly quickly so rapid drying (in a flow of air at 40 degrees C.) of at least some of a collection is a good idea. I would be happy to look at dried material. Usually fruits in limited quantities, so as many fruitbodies as possible at different stages of maturity. Please send to:

Derek Schafer 3 Keiths Wood Knebworth

SG3 6PU